

Nutritional benefits of flavocoxid in patients with osteoarthritis: efficacy and safety

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Abstract: Osteoarthritis (OA) is a major age-related, chronic disease in the United States that results in significant morbidity and functional loss as well as substantial financial outlay. Dietary habits have been shown to influence the metabolic and inflammatory processes involved in OA. For example, a lower intake of antioxidants is associated with increased incidence of OA while an increased intake of flavonoids and omega-3 polyunsaturated acids decreases production of inflammatory mediators and the incidence of OA. These observations argue that dietary habits contribute to the metabolic and inflammatory etiology of chronic diseases such as OA. Flavocoxid is a botanically-based formula composed of concentrated and standardized flavonoids from the botanicals *Scutellaria baicalensis* and *Acacia catechu*. Flavonoids constitute a large class of compounds known as polyphenols, ubiquitous in plants and consumed regularly in the human diet. Flavocoxid was developed by extensive high throughput screening and enzymatic testing and two specific flavonoids, catechin and baicalin, were identified that possess potent anti-inflammatory properties. Flavocoxid acts by managing both the cyclooxygenase and 5-lipoxygenase pathways of arachidonic acid metabolism thought to underlie the pathological processes of OA. Preclinical and clinical evidence suggests that flavocoxid provides effective and safe dietary management of the underlying metabolic processes of OA.

Keywords: cyclooxygenase, 5-lipoxygenase, inflammation, flavonoids

Introduction

Humans have sought and used botanical products for medicinal purposes for thousands of years. Many of the pharmaceuticals in use today are purified extracts or derivatives of substances of botanical origin.¹ Ethnopharmacology has become a major scientific specialty both academically and commercially with a number of pharmaceutical companies devoting a portion of their research and development budgets to exploration in areas such as the Amazon basin, Africa, the South Pacific, and East and Southeast Asia, generally with teams of ethnobotanists, biochemists, pharmacologists, and anthropologists. Many compounds have been “discovered” that have biologic activity on almost all physiologic systems and processes from cancer to ulcers, headaches to arthritis, goiters to heart function.^{2,3} Among these, flavonoids, with more than 9,000 molecular species described, have been found to have activity in many physiological systems. With the rapid extinction or absorption of indigenous peoples, it is feared that much traditional, orally transmitted, therapeutic knowledge will be lost thereby making the discovery of botanically-based therapeutic molecules more difficult by requiring more “random” sampling of botanical sources.

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Though flavonoid molecules have been known for decades to possess anti-inflammatory activity, only recently have any of these molecules entered OA clinical trials and found their way to market. This report will focus on the detailed preclinical and clinical investigation of flavocoxid, an anti-inflammatory medical food composition marketed for the metabolic management of OA under the brand name Limbrel®. Other flavonoid-based formulations have been tested in small, pilot clinical studies. For example, flavonoid extracts from *Garcinia kola* were equivalent to naproxen and celecoxib and better than placebo in addressing the pain, stiffness, and mobility of OA patients ($n = 21$)⁴ and, in pilot, randomized double-blind studies, pine bark extracts (Pycnogenol®) improved Western Ontario and McMaster University Osteoarthritis Index (WOMAC) composite scores for pain, stiffness, and mobility compared to the placebo control groups.^{5–7} Limbrel, however, is the only flavonoid, anti-inflammatory therapeutic currently marketed in the United States as a prescription therapeutic for OA.

OA: prevalence and costs

OA is the leading cause of musculoskeletal morbidity and functional loss in humans, its effects increasing with advancing age.^{8–10} In the US, it is estimated that 25% of people age 55 and over or as many as 40 million people have knee pain, and at least half of this population has had a radiographic diagnosis of OA.^{11,12} The prevalence of OA is expected to increase to 60 million by the year 2020 as the population ages.¹³ In 2001, the estimated prevalence of self-reported chronic joint pain was 33% of adults 18 and older, of which 10% carried a medical diagnosis of arthritis. Although the type of joint disease was not specified in this survey, OA was certainly the major component.¹³ Data from the National Center for Health Statistics 2006 Health Interview Survey reveals that 25% of adults aged over 18 years had chronic joint symptoms and 21% of adults were told by a health professional that they had arthritis. These statistics are almost identical to British data reported by Peat et al.¹¹ Not surprisingly, while only 6.9% of people aged 18–44 years carried a diagnosis of arthritis, the incidence increased to 28.7% between the ages of 45–64 years, 48% between 65–74 years, and remained relatively constant at 51.3% above 75 years.¹⁴ OA is a major cause of impaired mobility and expected to be the fourth leading cause of disability in the US and Western Europe by the year 2020.^{8,15,16} This has significant societal and financial implications. By one recent estimate, the average annual costs per person in an insured population of OA patients was \$8,601 for direct medical costs including treatment of drug induced side effects, \$2,941 for

drugs, and \$4,603 for indirect costs such as time lost from work.¹⁷ These data are remarkably consistent across time and populations.^{9,18–21} Reginster²² estimated the cost of all forms of arthritis at between 1%–2.5% of GNP for all Western nations and noted that poor quality of life issues related to arthritis exceeded those for gastrointestinal, respiratory, and cardiovascular diseases.²²

When evaluating the results of clinical, epidemiological, or socio-economic studies, it is important to understand the population under consideration, since approximately 50% of adults aged over 55 years with knee pain do not have radiographic evidence of OA and an equal percentage with radiographic OA do not have pain.^{11,23} Furthermore, when considering symptomatic, structural, or functional significance of disease or therapeutic intervention, the pattern of joint involvement must be defined. For example, aside from the concept of pain, impairment of mobility from OA of the hip or knee raises entirely different quality of life (QOL) and activities of daily living (ADL) issues than those arising from the impairment of fine motor function that accompanies moderate-severe hand OA.

Metabolic considerations in OA

The etiology of OA is multifactorial including things such as trauma, inflammation, obesity, and genetic predisposition. It is only relatively recently that, with a substantial increase in our understanding of the biochemical processes involved in the degradation and repair of articular cartilage and subchondral bone, the etiologic roles of nutritional factors and lifestyle have become more appreciated.

Much of the tissue injury and symptoms of pain and stiffness has been demonstrated to be related to products generated by metabolism of arachidonic acid (AA) derived from dietary intake and of cell membrane phospholipids by the action of phospholipase-A2. Conversion of AA to pro-inflammatory mediators proceeds by parallel enzymatic and oxidative pathways (Figure 1).^{24,25}

Production of prostaglandins (PGI₂, PGE₂, PGF₂) and leukotrienes, particularly LTB₄, LTC₄, and LTD₄, the end products of the cyclooxygenase (COX-1, COX-2) and 5-lipoxygenase (5-LOX) systems, respectively, results in recruitment of inflammatory cells and fluid as well as induction of cytokines, metalloproteinases, and other agents that contribute to cartilage breakdown and impairment of normal reparative processes.²⁵ An imbalance of thromboxanes and prostacyclins (from relatively excessive COX-1 or COX-2 inhibition, respectively) may account for some or most of the antiplatelet and cardiovascular toxicity seen with these agents, primarily caused by vasoconstriction of arteries, ultimately leading

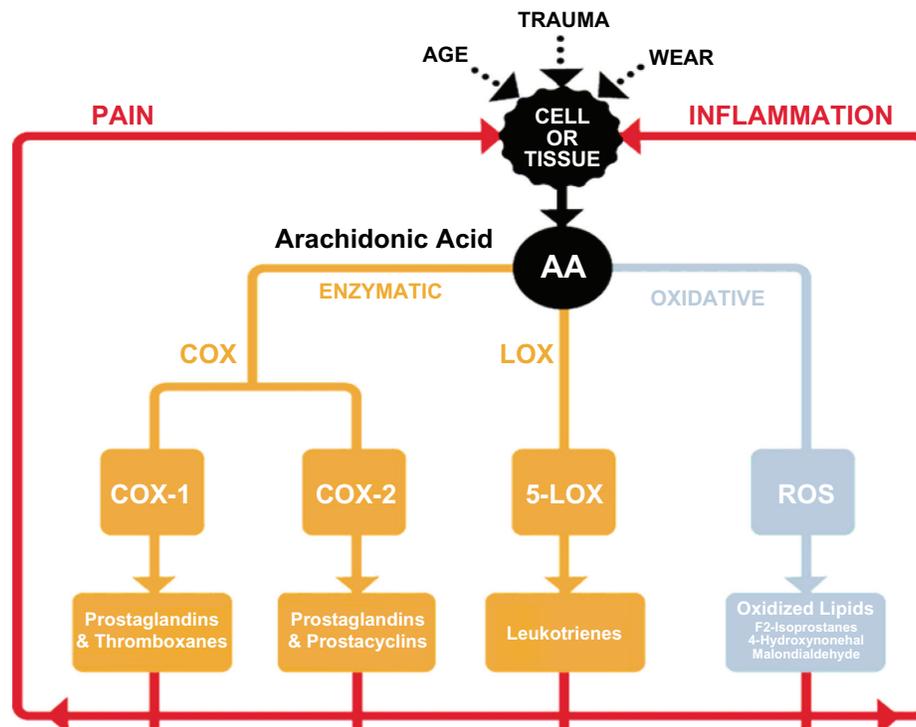


Figure 1 Metabolic pathways of arachidonic acid metabolism. AA is enzymatically metabolized to various prostaglandins, thromboxanes, prostacyclins, and leukotrienes through the COX-1, COX-2, and 5-LOX pathways. AA can also be oxidatively metabolized through the action of ROS to form inflammatory lipids such as isoprostanes, which are known to be elevated in the joints of OA patients

Abbreviations: AA, arachidonic acid; COX, cyclooxygenase; 5-LOX, 5-lipoxygenase; ROS, reactive oxygen species.

to edema, stroke, and myocardial infarction.²⁶ Different nonsteroidal anti-inflammatory drugs (NSAIDs) are either more COX-1 or COX-2 selective or relatively balanced on these two pathways.²⁷ For example, rofecoxib and valdecoxib are highly inhibitory for the cyclooxygenase activity of COX-2, while indomethacin, flurbiprofen, and ketorolac are highly inhibitory for the cyclooxygenase activity of COX-1. Sodium salicylate, due to its inherent mechanism of action, is a relatively balanced inhibitor of cyclooxygenase activity for COX-1 and COX-2.²⁷

In the VIGOR trial, rofecoxib showed a four fold increase in cardiovascular events (heart attack, stroke, and blood pressure changes) compared with naproxen.²⁸ A retrospective study of over 54,000 patients in two different health plan analyses demonstrated a three fold increase in heart attack over placebo among elderly adults in the first 90 days when taking >25 mg per day of rofecoxib.²⁹ Similarly, valdecoxib showed cardiovascular complications after cardiac surgery and in retrospective analysis of several clinical trials.³⁰ Finally, celecoxib has been shown to increase heart attacks, stroke, and other cardiovascular complications in a dose dependent manner (2.3% at 200 mg per day, 3.4% at 400 mg per day).³¹ Rofecoxib was voluntarily withdrawn from the market in late 2004 and valdecoxib was withdrawn in early 2005. In February 2005, a Food and Drug

Administration (FDA) advisory committee did not recommend that celecoxib be removed from the market, but recommended new “black box” warnings for all NSAIDs including selective COX-2 inhibitors.

Selective inhibition of COX-2 was once thought to be a way to reduce the incidence of gastric ulceration.^{28,32,33} Chronic use of these products, however, was shown to lead to a high incidence of ulceration.³⁴ Understanding AA metabolism and the importance of balancing COX-1 and COX-2 enzyme activity has become central in prescribing decisions for traditional NSAIDs, selective inhibitors, or recommending alternative therapies. Cyclooxygenases represent only one set of metabolic pathways to consider. Other inducible factors, such as cytokines and alternate inflammatory pathways like 5-LOX, also contribute to the pathogenesis of OA.

Evidence suggests that significant mechanical strain in a joint results in up-regulation of the pro-inflammatory cytokines TNF α and IL-1 β .^{35,36} IL-1 β then triggers production of reactive oxygen species (ROS) that induce expression of the pro-inflammatory transcription factor, nuclear factor kappa-B (NF- κ B),^{37,38} triggering an inflammatory cascade that, in turn, leads to up-regulation of COX-2³⁹ and chondrodestructive metalloproteinases.⁴⁰ NF- κ B is an important transcription factor complex that regulates the expression of more than

400 genes involved in immune function and inflammation.^{41,42} In an inactivated state, NF- κ B is bound by the inhibitory protein I κ B α within the cytosol.⁴³ Upon activation by extracellular signals and ubiquitination of I κ B α , free NF- κ B rapidly translocates to the nucleus where it activates the transcription of target genes including those encoding pro-inflammatory cytokines, adhesion molecules, chemokines, and inducible enzymes such as COX-2, 5-LOX, and inducible nitric oxide synthase (iNOS).⁴⁴ This activation and induction of inflammatory factors leads directly to increases in tissue destruction in OA and other chronic diseases. Modulation of NF- κ B is a key element in managing the metabolic aspects of joint deterioration.

In the past decade, several agents that inhibit both COX and 5-LOX pathways have entered clinical development. Many of these so-called “dual inhibitors” do not cause some of the side effects that are typical of NSAIDs or COX-2 inhibitors, including the cardiovascular, renal, and gastrointestinal damage and discomfort caused by traditional NSAIDs.⁴⁵ For example, it has been suggested that NSAID induced gastric inflammation is largely due to metabolites of 5-LOX, particularly LTB₄.⁴⁶ Leukotrienes, and especially LTB₄, are found in high concentrations in the walls of NSAID induced gastric ulcers and represent the primary AA metabolites within the gastric mucosa following prostanoid inhibition. Their presence suggests a shunt of AA metabolism toward leukotriene production as a result of COX inhibition.⁴⁷ These compounds are highly leukoattractive and contribute to a significant amount of the NSAID induced gastric epithelial injury.⁴⁸ Additional evidence for 5-LOX shunting caused by NSAIDs can be seen in the production of leukotrienes in patients on aspirin or similar cross reacting NSAIDs who develop asthmatic symptoms,^{49,50} in animal models of NSAID induced cardiac ischemia,⁵¹ and human OA synovial explants exposed to NSAIDs.⁵² Taken together, these characteristics suggest that there may be distinct advantages to dual inhibition of COX and 5-LOX over COX-1 and/or COX-2 alone with regard to both increased efficacy and improved safety profile.

Role of nutrition in OA

Epidemiological studies have shown that Asian and Mediterranean countries have a substantially lower incidence of knee OA suggesting an etiologic role for nutritional factors.¹⁵ Excess consumption of omega-6 fatty acids may account, in part, for this difference. Arachidonic acid is derived from the dietary intake of AA itself and essential fatty acids (EFA), linoleic acid, and α -linolenic acid followed by sequential desaturation and elongation.⁵³ Fatty acid imbalances are commonly seen in patients with chronic inflammatory conditions such as arthritis. Osteoarthritic joints typically show increased levels of lipid and AA accumulation which correlate with histological

severity.^{54,55} Fatty acid levels in subchondral bone have been shown to be 50%–90% higher in OA patients compared to controls.⁵⁶ Dietary lipids have also been shown to modify the fatty acid composition of cartilage.⁵⁷ In addition, clinical studies have shown a strong linkage between metabolic defects in EFA metabolism or an overabundance of fatty acids that lead to OA.⁵⁸

Flavonoids and other dietary antioxidants have been shown to slow the progression of OA,⁵⁹ possibly by managing the production of inflammatory oxidized lipids, enzymatically formed AA metabolites and reduction of transcriptional factors which induce other inflammatory components. This metabolic modulation may restore a more normal balance of fatty acid metabolism in the presence of excess omega-6 and AA dietary intake. Medical foods are ideally positioned to provide the clinical dietary management of chronic diseases like OA.

Regulatory considerations

Unlike drugs, medical foods are not developed under Investigational New Drug (IND) procedures or licensed under New Drug Application (NDA) requirements, but are subject to a specific set of governing regulations of their own. They are not “approved” by the FDA in the same manner as drugs, but are required to support all claims with good laboratory and clinical science. The category of Medical Foods was created in an addendum to the Orphan Drug Act of 1988. Prior to this they were regulated as drugs. The definition of a medical food was restated in the FDA’s Final Rule on Mandatory Nutritional Labeling, January, 1993. Specifically, a product classified as a medical food must meet the following requirements, among others:

- It is a specially formulated and processed product (as opposed to a naturally occurring foodstuff used in its natural state).
- It must be consumed or administered enterally, either by ingestion or intragastric tube.
- It provides nutritional support specifically modified for the management of the unique nutrient needs that result from the specific disease or condition, as determined by medical evaluation.
- The nutritional need could not reasonably be supplied by dietary alteration alone.
- It is intended to be used under medical supervision (ie, by prescription).
- It is intended only for a subject receiving active and ongoing medical supervision wherein the subject requires medical care on a recurring basis, ie, medical foods are intended for a diseased population.
- It is composed of ingredients designated as generally recognized as safe (GRAS). For an ingredient to achieve

GRAS status requires not only a technical demonstration of non-toxicity, but also an agreement of that safety after extensive peer review by an independent panel of experts in the field.

- It is based on recognized scientific principles.

Medical foods vs dietary supplements

The terms medical food and dietary supplement are not interchangeable.⁶⁰ These are discrete regulatory classifications with specific statutory definitions for each. The use of medical foods, even though they are composed of GRAS ingredients, must be prescribed and monitored by a physician while supplements are available over the counter. Moreover, supplements cannot make disease claims as their intended use is to preserve health in healthy people while medical foods, because they are intended for use in a diseased population, must make disease claims.

Flavocoxid

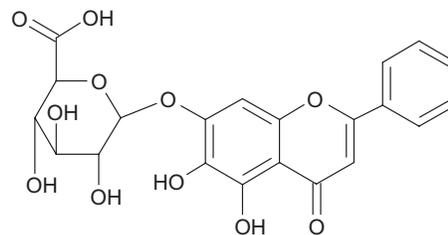
Flavocoxid is classified as a medical food under US FDA regulations. The product is a proprietary blend of processed and standardized free-B-ring flavonoids (baicalin; Figure 2a) derived from the plant *Scutellaria baicalensis* and flavans (catechin; Figure 2b) derived from the plant *Acacia catechu*.

As outlined in the following sections, flavonoids, in this specific combination, demonstrate a high degree of inhibition for COX-1, COX-2, and 5-LOX as well as other anti-inflammatory properties stemming from a strong antioxidant capacity. Flavocoxid's ingredient specification and analytical methods follow prescription drug standards and the product is manufactured according to current FDA Good Manufacturing Practices (GMP) for prescription drugs. This is a higher standard than that required for over the counter drugs, foods, or dietary supplements.

In vitro studies: anti-inflammatory properties

To isolate the components of flavocoxid (also known by the laboratory code UP446 in earlier publications), *in vitro* high throughput and enzyme assays, using recombinant ovine isozymes, were used to measure peroxidase (POX) inhibition for both COX-1 and COX-2 of more than 2,500 plant extracts.⁶¹ For isolation and characterization of the 5-LOX inhibition activity, a plant-based 5-LOX enzyme was used since no human isoform was commercially available at the time. Candidate extracts were then further purified by high performance liquid chromatography (HPLC) to single chemical entities, in this case, baicalein and catechin, to isolate

A. Baicalin (MW = 446.37)



B. Catechin (MW = 290.27)

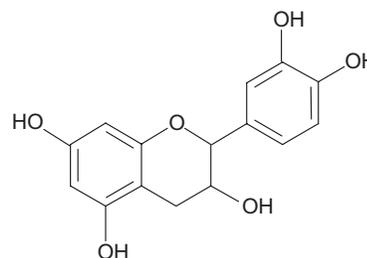


Figure 2 Molecular structure of baicalin and catechin components of flavocoxid.

the corresponding POX, for COX-1 and COX-2, and 5-LOX inhibitory activities.⁵⁹ Both enzymatic inhibition studies as well as cell-based assays were then utilized *in vitro* to examine the anti-inflammatory properties and mechanism of action of flavocoxid. To determine the IC₅₀ dose levels *in vitro* for POX activity inhibition within the COX enzymes, flavocoxid was titrated into the purified enzyme system. One could argue that inhibition of the POX activity is not a direct measure of COX inhibition since NSAIDs inhibit the COX activity of COX-1 and COX-2, rather than the POX activity. The COX activity of both enzymes first oxygenates AA to produce PGG₂, which then is converted to PGH₂ by the POX activity followed by cell-specific synthetase conversion to individual PGs or thromboxanes (TXs).⁶² Since the two enzyme activities are coupled and required to produce the end-product, inhibition of one or both activities reflects downregulation of overall COX enzyme activity and end product generation.⁶³

In the POX inhibition assay to determine the half maximal inhibitory concentration (IC₅₀) for COX-1 and COX-2, there was an exponential rise in inhibitory activity with flavocoxid doses between 10–100 µg/mL with nearly 100% inhibition being achieved at the higher dose levels. The IC₅₀ for both COX-1 POX and COX-2 POX inhibition was 15 µg/mL.⁶⁴ Further work is currently underway to elucidate the specific inhibitory capacity of flavocoxid for the COX moiety within both COX-1 and COX-2 using a more traditional oxygen consumption assay.^{65,66} Although the complete mechanism of action for

inhibition of the COX enzymes has not been elucidated, the 5-LOX inhibitory activity has been determined.

The 5-LOX IC_{50} was also determined by titration of flavocoxid into this purified enzyme system using a commercial 5-LOX screening kit.⁶⁴ Again, there was a steep rise in inhibitory activity between 10–100 $\mu\text{g/mL}$ concentrations of flavocoxid with an IC_{50} of 25 $\mu\text{g/mL}$ for 5-LOX inhibition. Enzyme assays represent a useful tool for elucidating mechanism of action, but measurement of the end products is also necessary. Cell assay are the principle tool which has been used to verify the findings from the enzyme assays.

Burnett et al found that PGE_2 levels from COX-2 were reduced as flavocoxid was titrated into human osteosarcoma (HOSC) cultures.⁶⁴ Celecoxib was a much better COX-2 inhibitor in this assay, perhaps reflecting its specific COX inhibitory activity compared to the POX inhibition of flavocoxid. Flavocoxid also inhibited the production of LTB_4 in THP-1 monocyte cells when titrated into culture.⁶⁴ In another set of cell assays performed in rat peritoneal macrophages induced to an inflammatory phenotype with lipopolysaccharide (LPS), flavocoxid significantly blunted NF- κB activation and restored $\text{I}\kappa\text{B}\alpha$ expression, the cytoplasmic modulatory factor for NF- κB .⁶⁷ As a consequence of NF- κB modulation, COX-2, 5-LOX, iNOS, and TNF α protein expression was reduced when flavocoxid was titrated into culture. The metabolites of COX-2, 5-LOX, and iNOS, PGE_2 , LTB_4 , and nitrate were similarly reduced. Finally, flavocoxid, acting as a direct antioxidant, also decreased the formation of malondialdehyde from AA.⁶⁷ It is this antioxidant capacity that is most likely responsible for the modulation of NF- κB activation.²⁵ Further work on the specific antioxidant capacity as well as genomic mechanisms involved in the modulation of inducible inflammatory factors is under way.

Mutagenicity

To understand the toxicity profile of flavocoxid, a number of *in vitro* assays including cytotoxicity,⁶⁸ AMES mutagenesis assay,^{69,70} and human liver microsome CYP450 drug interaction assays⁷¹ were performed. In a cytotoxicity study in THP-1 monocytes, flavocoxid, aspirin, and ibuprofen caused a 5%–10% cell death while indomethacin and celecoxib resulted in 80%–100% cell mortality when titrated into culture between 50–100 $\mu\text{g/mL}$.⁷² In contrast, flavocoxid displayed low cytotoxicity at the highest concentration tested, 100 $\mu\text{g/mL}$. In addition, flavocoxid caused no mutagenicity as judged by a reduction of the *Salmonella typhimurium* bacterial background lawn or decrease in the number of spontaneous revertants at any dose level and in any of the

five different strains tested. In the presence of liver enzymes for metabolic activation, flavocoxid did not increase the incidence of revertants, but was slightly cytotoxic in three of five strains of *S. typhimurium*.

Drug interactions

Compounds that inhibit the CYP450 isoenzymes by 50%, or more, may significantly affect metabolism of one or more drugs and should be further characterized by determination of an IC_{50} and a pharmacokinetic study in humans to assess the maximum safe *in vivo* exposure. Flavocoxid at a concentration of approximately 10 μM showed moderate inhibition (23%) of CYP1A2 and only a slight inhibition of CYP3A4 (11%),⁷² the principal metabolizing enzyme of 50% of all pharmaceutical drugs.^{73,74} Similar inhibition findings were noted for CYP2C9, CYP2C19, and CYP2D6 (11%–16%). Because flavocoxid did not inhibit any one of the five principle drug metabolizing enzymes to 50%, it does not have major drug interactions and should be safe to take with most medications.

These accumulated *in vitro* studies suggest that flavocoxid acts via multiple inflammatory pathways on the enzymatic, antioxidant, gene, and protein expression levels and may be safe in patients due to low cytotoxicity, lack of a mutagenic effect, and absence of major CYP450 drug metabolizing inhibitions. The proof of any good therapeutic effect for a specific condition or disease, however, is the effect seen *in vivo*. Therefore, flavocoxid has been tested in both preclinical and clinical studies to evaluate safety and efficacy.

Preclinical: safety studies

Acute (14 days) and subchronic (91 days) toxicity studies were performed⁷² in 5- to 6-week old male and female imprinting control region (ICR) mice by feeding 250, 1,250, 2,500, and 10,000 mg daily human equivalent dosages of flavocoxid.⁷⁵ No differences in weight, gross pathology, hematology, or blood chemistry and histology of liver, kidney, duodenum, and stomach tissues were found in the treated vs untreated animals. A similar study was performed in 6- and 18-month old F344 male rats fed 0 or 34 mg/kg/day of flavocoxid for 9–10-weeks and, again, no abnormalities in weight, gross pathology, blood counts, or chemistry or histological examinations were found.⁷² The lack of tissue damage in the duodenum or stomach tissues is particularly significant in rats since this species is sensitive to the adverse effects of traditional NSAIDs, forming gastrointestinal lesions when dosed with as little ibuprofen as 2 mg/kg/day, equivalent to approximately 140 mg/day in humans.⁷⁶ Other flavonoids have

also been shown to either be gastric protective or have been used for the treatment of gastric ulcerations.⁷⁷

Anti-inflammatory efficacy models

Anti-inflammatory efficacy was determined initially in animals by using AA-based topical ear and intra-articular ankle swelling assays.⁶⁴ Arachidonic acid has been shown to induce the production of prostaglandins and leukotrienes when applied to various tissues.⁷⁸ The presence of both COX and LOX inhibitors has also been shown to abate swelling responses in AA-induced inflammatory models.⁷⁹ Flavocoxid (50, 100 or 200 mg/kg), administered by oral gavage to 4–5-week old ICR mice (Harlan Labs) 12 hours before AA application to ears or injection into ankles, blocked AA-induced inflammatory effects.⁶⁴ Indomethacin (50 mg/kg) gave a comparable response in the same assay. In the intra-articular AA model, there was a corresponding restoration of function as determined by comparing the flavocoxid treated and nontreated animals' abilities to run on a rotating rod.⁶⁴ More direct models of measuring flavocoxid's efficacy have also been used in animals with OA.

A formulation of flavocoxid specifically for canines was compared to a glucosamine hydrochloride/chondroitin sulfate/manganese ascorbate formulation in a multisite, double-blind, randomized, direct-comparator trial in dogs with OA weighing at least 15 lbs.⁸⁰ The flavocoxid formulation (n = 33) showed statistically significant improvement in discomfort over the combination formulation of glucosamine hydrochloride/chondroitin sulfate/manganese ascorbate formulation (n = 36) using veterinarian and owner visual analog scale (VAS) assessments. At both the interim (28 days) and final analysis (56 days), the flavocoxid formulation was more than twice as effective as the glucosamine hydrochloride/chondroitin sulfate/manganese ascorbate formulation at managing discomfort in this canine population. The VAS for discomfort was shown to be statistically separable at 56 days for flavocoxid formulation over the glucosamine hydrochloride/chondroitin sulfate/manganese ascorbate formulation. Adverse events were generally mild and equivalent in both groups.

Clinical studies: pharmacokinetic study

Baicalin and other flavonoids are partially water-soluble molecules and are absorbed intact to varying degrees in the gastrointestinal tract following oral administration via a glucose transporter system such as GLUT-4. The vast majority of the glycosidic form resists digestion by acid in the stomach and arrives intact in the duodenum.⁸¹ The tissue and

species-specific glycosidases, present in the gastrointestinal microflora and brush border membrane, convert the flavonoid glycosides into aglycones such as baicalein.^{82–84} A small percentage (<10%) of the aglycone baicalein is restored to baicalin in the liver following absorption, and circulates in the body primarily bound to plasma proteins.^{85,86} Aglycones and other flavonoids are often further metabolized by C-ring fission, resulting in the formation of two phenolic products: a methylation and a sulfation product.^{87,88} The proportion of each modification seems to vary according to the type and amount of flavonoid ingested, species, sex, and food deprivation with glucuronidation favored over sulfation and sulfation favored over methylation.^{89,90} The conversion of baicalin to baicalein is highly variable in each individual depending on the concentration and type of intestinal flora in the small intestine.⁹¹ Excretion occurs primarily via the biliary system.

Catechin (also known as (+)-catechin) is readily absorbed in the small intestine and does not undergo any predigestion.⁹² The absorption of catechin from the gastrointestinal tract is rapid and immediately followed by a conjugation step in the intestinal mucosa to produce a methylated metabolite.⁸⁸ The glucuronidated catechins enter the portal vein and are either sulfated in the liver, or sulfated and methylated in the kidneys. Excretion occurs in the bile and urine.⁹³ Animal studies have shown that the majority of catechin is excreted in the urine and in the breath, as respired carbon dioxide.⁹⁴ The results of this study, in which volunteers were administered (4×) 500 mg of (+)-catechin, indicated that more than half of the original dose was absorbed as the intact flavanol from the human intestine.⁹⁵ The half-life for catechins has been reported to range from 2–3 hours⁹⁶ and plasma levels are observed to peak ~ 3 hours after administration in humans, with levels still detectable 120 hours later, probably due to reabsorption and enterohepatic circulation.⁹⁵ Overall, the hepatic extraction of orally administered catechin is rapid and efficient, with relatively low levels of the unchanged compound present in the plasma. Since the relative pharmacokinetic profile of catechin is known, and it exists as a minority of the composition in flavocoxid, a pharmacokinetic study was undertaken with a single dose of flavocoxid in fasted subjects to determine the baicalin absorption and excretion profile.

Ten fasted human subjects received 500 mg of flavocoxid under controlled conditions. Plasma samples were collected at ½, 1, 2, 4, 8, and 24 hours and again on day 7 post-dose. After digestion of the plasma samples with β-glucuronidase and sulfatase, serum baicalein levels were measured and pharmacological parameters including area under curve (AUC), maximum concentration (C_{max}), time to reach C_{max}

(T_{max}), and half-life ($T_{1/2}$) determined. Although considerable individual variation was noted in the data, the average AUC was 7,007 $\mu\text{g}/\text{mL}/\text{hour}$, C_{max} was 0.93 $\mu\text{g}/\text{mL}$, T_{max} was 5.8 hours, and the $T_{1/2}$ was approximately 11–12 hours. The average pharmacokinetic profile over the first 24 hours is shown in Figure 3.

This study represents a single dose pharmacokinetic profile. A steady-state pharmacokinetic study may be needed to elucidate the absorption and excretion profile of baicalin and catechin in combination. In addition, not all of the metabolite input can be accounted for when administering flavonoids orally, so further work may be needed to find the reservoirs in which these molecules reside.

Baicalein is primarily converted to 6-O-beta-glucopyranuronoside (M1), 6-O-beta-methyl-baicalein 7-O-beta-glucopyranuronoside (M2), baicalein 7-O-glucopyranuronoside (M3), 6-O-beta-glucopyranuronosyl-baicalein 7-O-sulfate (M4), and baicalein 6,7-di-O-beta-glucopyranuronoside (M5).⁹⁷ The M4 and M5 metabolites are the vast majority found in biliary secretion at about 40%–50% (of the 10% that reaches the liver) of the total input over 30 hours in a rat study whereas renal secretion of these metabolites accounts for under 10%.⁹⁸ Therefore, approximately 40% of all input flavonoid cannot be accounted for using these analyses. Some may show up in the feces, however, due to extensive degradation of baicalein by colonic microflora, exact quantification is difficult.⁹⁹ There may also be other reservoirs that account for missing flavonoids in these studies.

Several recent studies have demonstrated that the bioavailability of flavonoids may be much greater than initially believed.¹⁰⁰ It has been shown that flavonoid molecules bound by serum albumin are taken up by red blood cells (RBCs), which essentially act as a reservoir for these

molecules.^{101,102} Flavonoids are subsequently released from the RBCs when the albumin molecule becomes “desaturated” by the release of bound endogenous or exogenous substances. Therefore, it is reasonable to assume that the concentration of circulating flavonoids such as baicalein and catechin in human plasma is even greater when the amount associated with RBCs is considered. Furthermore, the relatively slow elimination half-life of baicalein of ~6 hours can be explained by its high affinity for albumin. About 90% of baicalin¹⁰³ and 45% of catechin¹⁰⁴ are bound by serum albumin. Alternatively, the slow release from RBC reservoirs may also explain the observed half-life of baicalein in our study. Other health factors may also affect flavocoxid bioavailability.

Two animal studies have suggested that antibiotics can reduce the level of glucoside processing microflora in the small intestine, thereby decreasing the conversion of baicalin to baicalein, and ultimately the uptake of baicalein into the blood.^{105,106} Therefore, the efficacy of flavocoxid may be affected while a patient is on a short course of antibiotics although this has not been confirmed in human studies.

Clinical studies

The flavocoxid composition has been studied in several controlled clinical trials. In an initial safety study, Burnett et al administered flavocoxid 250 mg or placebo every day to healthy volunteers ($n = 80$) for 5 weeks.⁷² Subjects were sex matched and recruited from ages 40 to 75 years. Subjects taking NSAIDs underwent a 2-week washout period before the baseline visit. Physical activity was not restricted and the subjects were not given any advice pertaining to diet. Safety was measured by the incidence of treatment-emergent adverse events (AEs) and laboratory abnormalities. Symptomatic AEs were few, mild, and comparable in treated and placebo groups. No mean values for plasma biochemistry of either group fell outside of the clinically acceptable ranges.

A 3-month safety study was performed in 59 OA subjects given flavocoxid 250 mg bid or placebo.⁶⁰ There were no differences in AEs or blood chemistries except for an apparent reduction in upper respiratory related AEs in the flavocoxid group (flavocoxid 4 vs placebo 17; $P = 0.0001$). Flavonoids, including catechins and flavonols, such as baicalin, have been studied for their antimicrobial activity and potential usefulness against infectious diseases in humans.^{107–109} Baicalin, purified from *S. baicalensis*, has been traditionally used as an anti-inflammatory and anti-allergic agent in the treatment of many inflammatory diseases ranging from bronchitis to atopic dermatitis.¹¹⁰ Gargling tea catechin extracts has been shown to significantly reduce the incidence of influenza

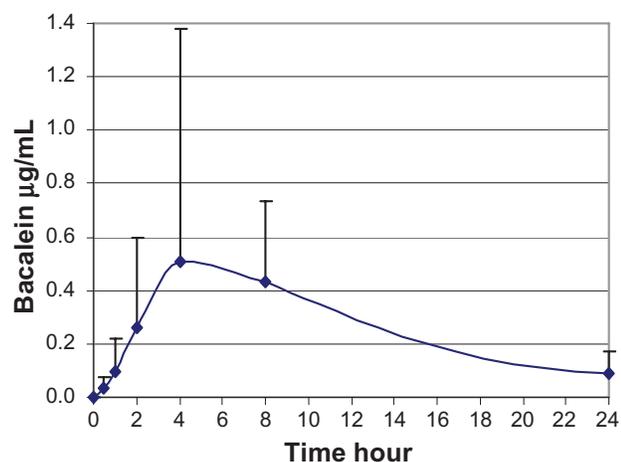


Figure 3 Pharmacokinetic profile of uptake and clearance of baicalein, the aglycone metabolite of baicalin.

infection in elderly nursing home residents compared to a control group receiving placebo.¹¹¹ This property of flavocoxid will be explored in more detail in future studies.

A 1-month randomized, multicenter, pilot efficacy study comparing flavocoxid 500 mg bid with naproxen 500 mg bid enrolled 103 subjects of whom 101 completed the study.¹¹² In this short-term study, flavocoxid and naproxen were statistically identical with regard to both efficacy (>80% response, overall) (Figure 4) and safety (~21% AEs, none serious).¹¹²

A 3-month randomized, double-blind, multicenter study comparing flavocoxid 500 mg bid (n = 106) with naproxen 500 mg bid (n = 114) has recently been completed (submitted for publication). There was no significant difference in efficacy measured by multiple conventional parameters including WOMAC scales, timed walk, and multiple VAS scales when the entire intent to treat population is considered. There was a statistically significant reduction in upper gastrointestinal AEs and edema in the flavocoxid group.

Flavocoxid has been marketed under the brand name Limbrel since May, 2004. Despite the vagaries of voluntary post-marketing reporting, there has been a notable lack of renal, cardiovascular, and upper gastrointestinal AEs. With more than 150,000 new prescriptions filled, the most common complaints are referable to lower gastrointestinal distress, primarily diarrhea, and flatulence. This is consistent with formal clinical trials where the incidence of these complaints has been about 5%, but has not resulted in any study subject drop outs. In clinical trials, elevated liver function tests have been noted, but have not been significantly different in frequency from naproxen. Occasional reports of allergic reactions such as rashes or hives (<0.2%) and rare cases (n = 6) of hypersensitivity pneumonitis requiring treatment with glucocorticoids and oxygen have been reported.

A multicenter, in-market rheumatological study in over 1,000 patients given 500 mg bid assessing response rate, AEs, gastrointestinal tolerability, and reduction in gastroprotective use has also been performed.¹¹³ Initial evaluation of results demonstrated that flavocoxid is well-tolerated in NSAID sensitive individuals, reduces the use of gastroprotectives, and has a high response rate. In addition, an anticoagulant study showing absence of flavocoxid effect on aspirin-induced blood thinning in an animal model, a platelet function study in patients taking 500 mg bid flavocoxid for 14 days (n = 10) showing no effect on bleed times or platelet aggregation and an open-label study in rheumatological practices of flavocoxid administered for 2 weeks at either 250 or 500 mg bid in warfarinized patients demonstrating no effect on international normalized ratio (INR) (n = 59) have been completed and

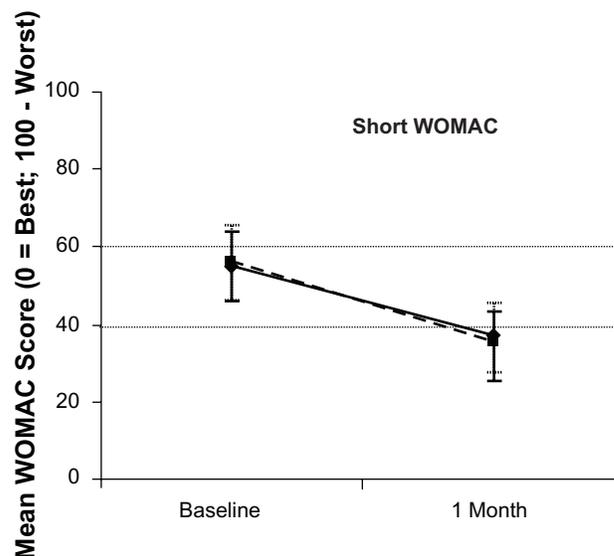


Figure 4 Improvements in efficacy with flavocoxid. Levy et al (2009) showed no statistically significant differences in WOMAC scores between the flavocoxid and naproxen arms during a 4-week clinical trial. The means and \pm SEM are shown for this efficacy measurement. Differences between groups were not significant ($P > 0.2$). Decreasing trends indicated improvement ($P \leq 0.001$). ♦ indicates flavocoxid, 500 mg bid ■ indicates naproxen 500 mg bid.

have been submitted for publication. These peer-reviewed trials will provide a better clinical profile for flavocoxid to aid clinicians in their prescribing decisions.

Conclusion

The goal of a metabolic therapy for OA is to reduce oxidative stress and restore a more normal balance of fatty acid metabolism in the presence of excess omega-6 and AA dietary intake. Flavonoids and dietary antioxidants, such as those contained in flavocoxid, have been shown to slow the progression of OA⁵⁹ possibly by managing the production of inflammatory oxidized lipids, enzymatically formed AA metabolites, and reduction of transcriptional factors which induce other inflammatory components. Further testing of the mechanism of action of flavocoxid is underway as well as testing to determine if flavocoxid is cartilage protective and/or restorative.

Flavocoxid (Limbrel), a prescription medical food based on a proprietary formula of purified flavonoid molecules, represents a new class of anti-inflammatory agents compared to traditional NSAIDs or COX-2 selective inhibitors. Specifically, flavocoxid acts as a balanced peroxidase inhibitor of COX-1 and COX-2 with additional 5-LOX inhibitory activity to avoid AA metabolic shunting toward leukotriene production. In addition, flavocoxid works via antioxidant mechanisms to downregulate the inducible production of inflammatory pathways, mRNA and proteins, and as a direct antioxidant to presumably decrease cartilage damage. Flavocoxid has also

been shown in two pilot safety trials to be safe vs placebo and in two efficacy studies to be equivalent to naproxen. As such, flavocoxid represents the first broad-based, flavonoid anti-inflammatory that has been well-studied for its mechanism of action, drug interactions, mutagenesis, and preclinical and clinical safety and efficacy to appear on the market. It represents a new option for the clinical dietary management of the metabolic aspects of OA.

Disclosure

The authors are employed by Primus Pharmaceuticals, Inc. who have licensed a compositional patent on flavocoxid (Limbril) and provided financial sponsorship for the associated clinical trials.

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